

IDH1 mutation detection in plasma circulating tumor DNA (ctDNA) and association with clinical response in patients with advanced intrahepatic cholangiocarcinoma (ICC) from the phase 3 ClarIDHy study

Elia Aguado-Fraile¹, Ghassan K Abou-Alfa^{2,3}, Andrew X Zhu^{4,5}, Teresa Macarulla⁶, Bin Fan¹, Parham Nejad¹, Sung Choe¹, Liewen Jiang¹, Camelia Gliser¹, Shuchi S Pandya¹, Bin Wu¹

¹Agios Pharmaceuticals, Inc., Cambridge, MA, USA; ²Memorial Sloan Kettering Cancer Center, New York, NY, USA; ³Weill Medical College at Cornell University, New York, NY, USA; ⁴Massachusetts General Hospital Cancer Center, Harvard Medical School, Boston, MA, USA; ⁵Jiahui International Cancer Center, Jiahui Health, Shanghai, China; ⁶Vall d'Hebron University Hospital, Vall d'Hebron Institute of Oncology (VHIO), Barcelona, Spain

Email: medinfo@agios.com

BACKGROUND

- Somatic mutations of isocitrate dehydrogenase 1 (*IDH1*) result in neomorphic catalytic activity, leading to the conversion of α-ketoglutarate to D(-)-2-hydroxyglutarate (2-HG)¹
 - Accumulation of 2-HG contributes to tumor initiation and progression through epigenetic dysregulation and a block in cellular differentiation^{2,4}
- Mutations in *IDH1* are commonly found in cholangiocarcinoma (CC), with higher incidence (~13%) among intrahepatic CC (ICC) cases⁵
- The mutant *IDH1* (m*IDH1*) inhibitor ivosidenib (IVO; AG-120) has been evaluated as monotherapy in CC across two clinical studies (phase 1 and phase 3)
- In the ongoing, global, phase 3 ClarIDHy study evaluating IVO vs placebo (PBO) in patients with nonresectable or metastatic m*IDH1*-CC (ClarIDHy, ClinicalTrials.gov NCT02989857):⁶
 - IVO demonstrated a favorable safety profile
 - Progression-free survival (PFS) for IVO was significantly improved relative to PBO, with a hazard ratio (HR) of 0.37 (95% CI 0.25, 0.54); p < 0.001
 - For IVO, 6-month and 12-month PFS rates were 32% (95% CI 23%, 42%) and 22% (95% CI 13%, 32%), respectively; no patients in the PBO group were progression free for > 6 months
- The emergence of actionable mutations in CC, including *IDH1*, highlights the relevance of molecular testing in this disease
 - Tissue-based genomic profiling remains the gold standard for personalized therapy in this indication; however, CC tumors are not easily accessible, and biopsies often yield suboptimal tumor cell content for genomic profiling⁷
- Previous work has demonstrated the feasibility of circulating tumor DNA (ctDNA) detection in patients with biliary tract cancer, including CC, and the mutational landscape of plasma appears similar to that of tissue, indicating that liquid biopsies are a reliable approach for genomic profiling at baseline as well as for disease monitoring upon treatment⁸⁻¹⁰
- Previous data from our group demonstrated the feasibility of m*IDH1* detection in plasma ctDNA from patients with CC enrolled in phase 1 studies of m*IDH1* inhibitors, with high concordance with m*IDH1* status in tumor tissue¹¹

OBJECTIVES

Baseline assessments

- To determine the concordance of m*IDH1* detection in plasma and formalin-fixed paraffin-embedded (FFPE) tumor tissue in a larger patient cohort
- To correlate plasma m*IDH1* variant allele frequency (VAF) with plasma 2-HG
- To explore the potential predictive value of m*IDH1* ctDNA levels for PFS

Longitudinal assessments

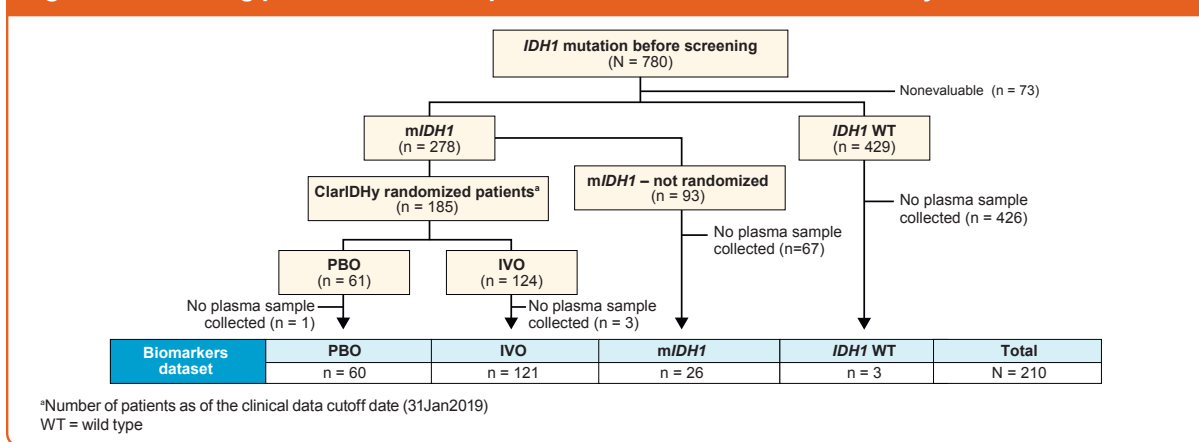
- To determine m*IDH1* ctDNA levels upon treatment with IVO or PBO
- To determine if *IDH1* mutation clearance in IVO-treated patients is associated with PFS

METHODS

- Archival FFPE tumor tissue samples were analyzed using OncoPrint Focus Assay for prospective central confirmation of m*IDH1* with 2.5% sensitivity. Tumor tissue samples were collected from 0.3 months up to 7.5 years before randomization (median 3.7 months)
- Pre-treatment plasma samples from all patients participating in screening were collected, and longitudinal samples from patients who were enrolled were obtained on Day (D) 1 of each treatment cycle. Blood samples were processed according to Sysmex plasma preparation instructions¹²
- BEAMing digital PCR (Sysmex) was used for the detection and quantification of five m*IDH1* alleles (R132C, R132H, R132L, R132S, and R132G) with 0.02% analytical sensitivity (0.04% for R132H)
- Baseline plasma levels of 2-HG were measured using a qualified liquid chromatography-tandem mass spectrometry method with a lower limit of quantitation of 30.0 ng/mL
- The clinical data cutoff date was 31Jan2019; for longitudinal m*IDH1* VAF analysis the biomarker data cutoff date was Mar2020

RESULTS

Figure 1. Screening plasma ctDNA sample collection flowchart and summary



RESULTS (CONTINUED)

Table 1. Concordance of m*IDH1* status in plasma and tissue

Detection in tissue, n	Positive	Detection in plasma, n	
		Positive	Negative
Positive	192	192	15
Negative	2	1	1

Screening m*IDH1* detection in plasma is highly concordant with mutations in tumor tissue

- The plasma ctDNA sample collection flowchart is shown in Figure 1
- Detection of m*IDH1* in plasma ctDNA was concordant with *IDH1* mutation status in tissue in 193 of 210 patients (92%) (Table 1)
 - 15 of 210 patients (7.1%) showed m*IDH1* detection in tissue but not in plasma
 - Tissue m*IDH1* VAF: median (range) 15.3% (3.8–37.0%)
 - Plasma m*IDH1* VAF: median (range) below detection limit (below detection limit, cutoff 0.02–0.04%)
 - 2 of 210 patients (0.95%) were deemed negative for m*IDH1* in tissue but showed detection in plasma
 - Tissue m*IDH1* VAF: 1.5% and 0.6% (tissue assay cutoff, 2.5%)
 - Plasma m*IDH1* VAF: 0.093% and 6.89% (plasma assay cutoff, 0.02–0.04%)
- The m*IDH1* allele detected was concordant, across all samples, with m*IDH1* detection in both tissue and plasma
- m*IDH1*-R132 allele distribution was similar between concordant and nonconcordant prescreening samples (Table 2)

Table 2. Similar m*IDH1*-R132 allele distribution in prescreening samples

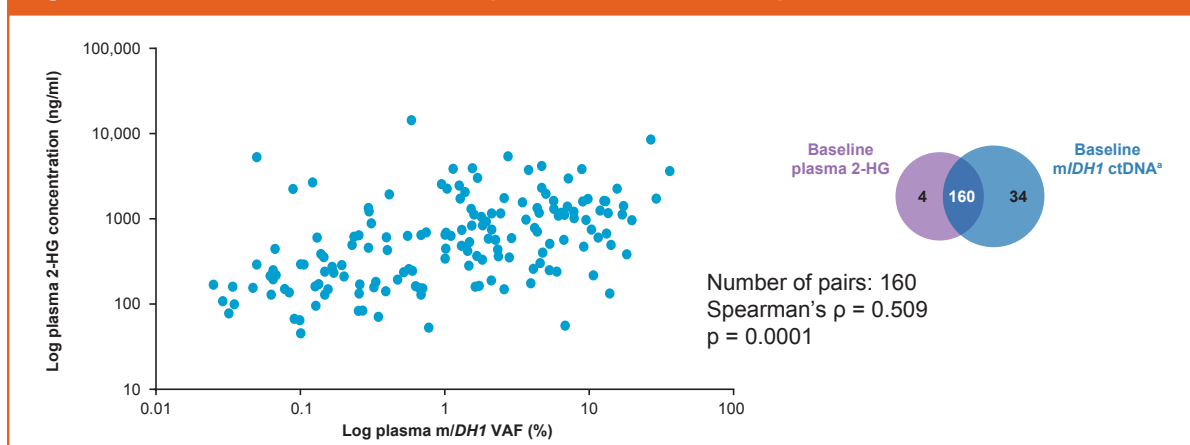
Allele	Concordant samples, n (%)	Nonconcordant samples, n (%)
R132C	135 (70.3)	13 (76.5)
R132L	29 (15.1)	3 (17.6)
R132G	22 (11.5)	1 (5.9)
R132H	2 (1)	0
R132S	4 (2.1)	0
Total	192	17

*Only m*IDH1* positive samples included

Baseline circulating m*IDH1* VAF correlates with plasma 2-HG levels

- Spearman's rank correlation analysis demonstrated a moderate correlation between plasma m*IDH1* VAF and plasma 2-HG (Figure 2)
 - This correlation was maintained when samples were separated by treatment arm:
 - IVO, Spearman's ρ = 0.57; p < 0.0001 (n = 105)
 - PBO, Spearman's ρ = 0.36; p = 0.006 (n = 55)

Figure 2. Correlation between baseline plasma m*IDH1* VAF and plasma 2-HG levels



*Patients without baseline m*IDH1* detection in plasma were excluded from this analysis (n = 16). 15 out of 16 were m*IDH1* positive in tissue

Lower levels of baseline m*IDH1* ctDNA are associated with longer PFS in IVO-treated patients

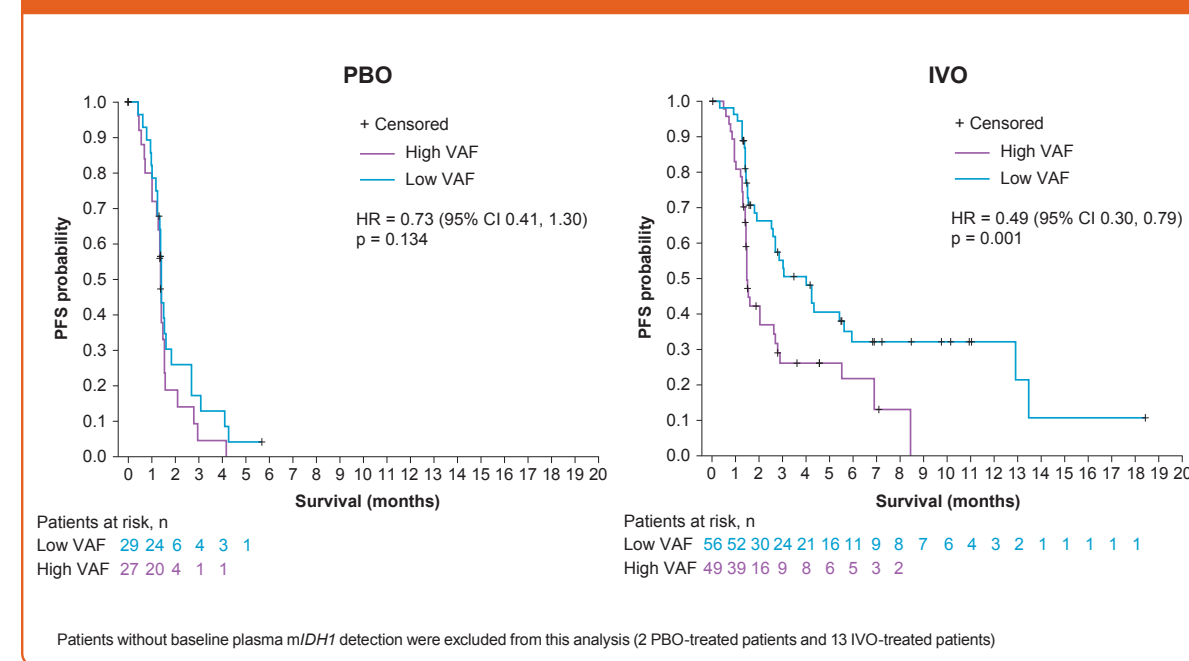
- Higher levels of baseline plasma m*IDH1* ctDNA were associated with shorter PFS in patients treated with IVO (Table 3, Figure 3)
 - No differences were found in the PBO control arm
 - Patients without baseline plasma m*IDH1* detection were excluded from this analysis (PBO, n = 2; IVO, n = 13) (Figure 1)
 - m*IDH1* VAF low vs high category was determined by calculating the median plasma VAF across all prescreening samples with m*IDH1* detection (n = 194)
 - VAF categories were defined as follows: low VAF, < 1.533; high VAF, ≥ 1.533
 - At screening, median VAF was similar in the PBO and IVO arms: 1.53 (n = 58) vs 1.41 (n = 108), respectively (p = 0.554)

Table 3. Median PFS in patients with low and high baseline plasma m*IDH1* VAF levels

	PBO		IVO	
	Low VAF n = 27	High VAF n = 29	Low VAF n = 49	High VAF n = 56
Median PFS (95% CI)	1.4 (1.3, 1.6)	1.4 (1.2, 1.6)	4.0 (2.5, 5.6)	1.5 (1.4, 2.6)
HR (95% CI)	0.73 (0.41, 1.30)		0.49 (0.30, 0.79)	
p-value	0.134		0.001	

m*IDH1* VAF category was determined by calculating the median plasma VAF across all prescreening samples (n = 194). VAF categories were defined as follows: low VAF, < 1.533; high VAF, ≥ 1.533

Figure 3. Association between baseline m*IDH1* ctDNA and PFS



m*IDH1* clearance was observed in a subset of IVO-treated patients

- The sample collection flowchart for longitudinal assessments is shown in Figure 4
- m*IDH1* clearance in plasma was found in IVO-treated but not in PBO-treated patients
 - m*IDH1* clearance was defined as m*IDH1* VAF below the assay's sensitivity for at least one on-treatment timepoint
- In IVO-treated patients, m*IDH1* clearance in plasma was found in 10 of 36 (27.8%) patients with PFS ≥ 2.7 months and 1 of 55 (1.8%) patients with PFS < 2.7 months (Figure 5, Table 4)
 - IVO demonstrated an improvement in PFS vs PBO, with a median PFS of 2.7 months vs 1.4 months for IVO and PBO, respectively⁶
- Mutation clearance was achieved at Cycle (C) 3 D1 or earlier in seven of eleven (63.6%) patients
 - Early clearance might predict a favorable outcome
- Plasma m*IDH1* clearance in IVO-treated patients was associated with longer PFS (Figure 6)
 - In patients with clearance, median PFS was 12.9 months
 - In patients without clearance, median PFS was 2.6 months

Figure 4. ClarIDHy longitudinal plasma ctDNA sample collection flowchart and dataset summary

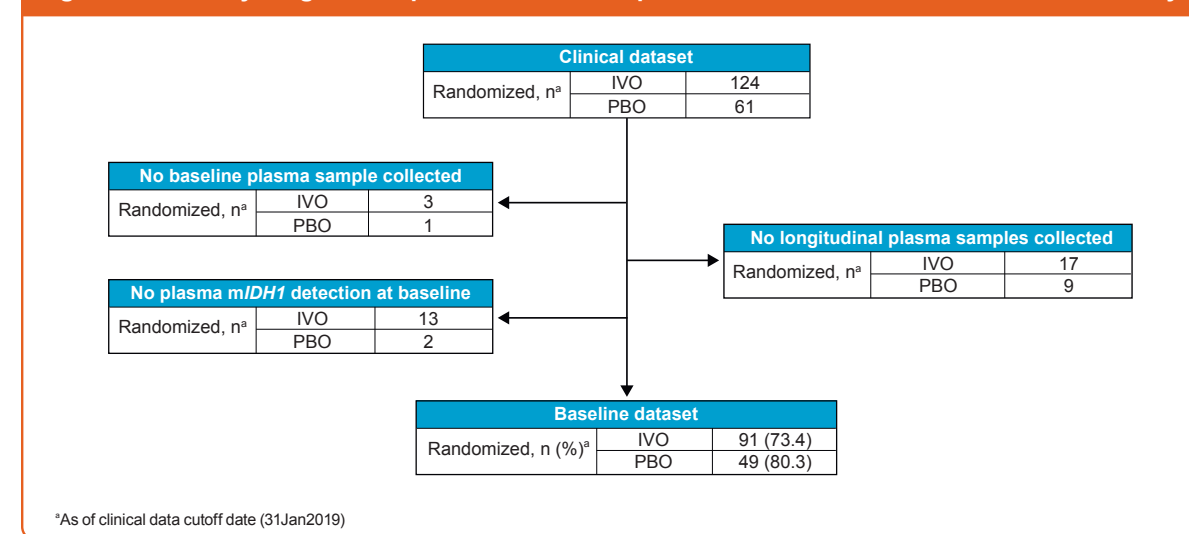


Figure 5. m*IDH1* clearance in plasma in a subset of IVO-treated patients

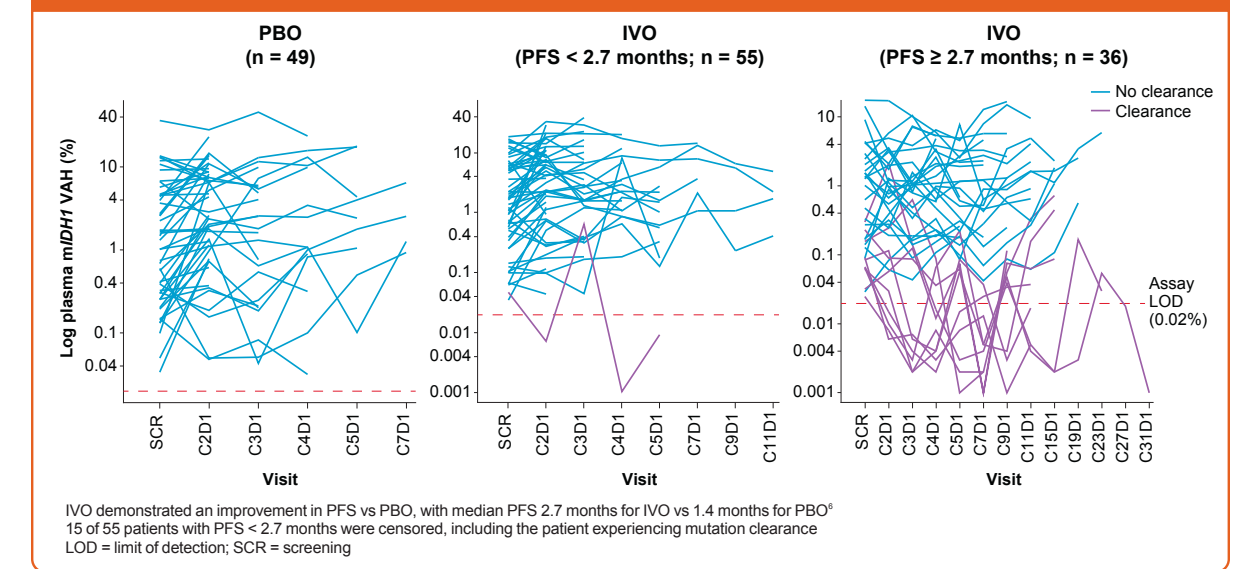
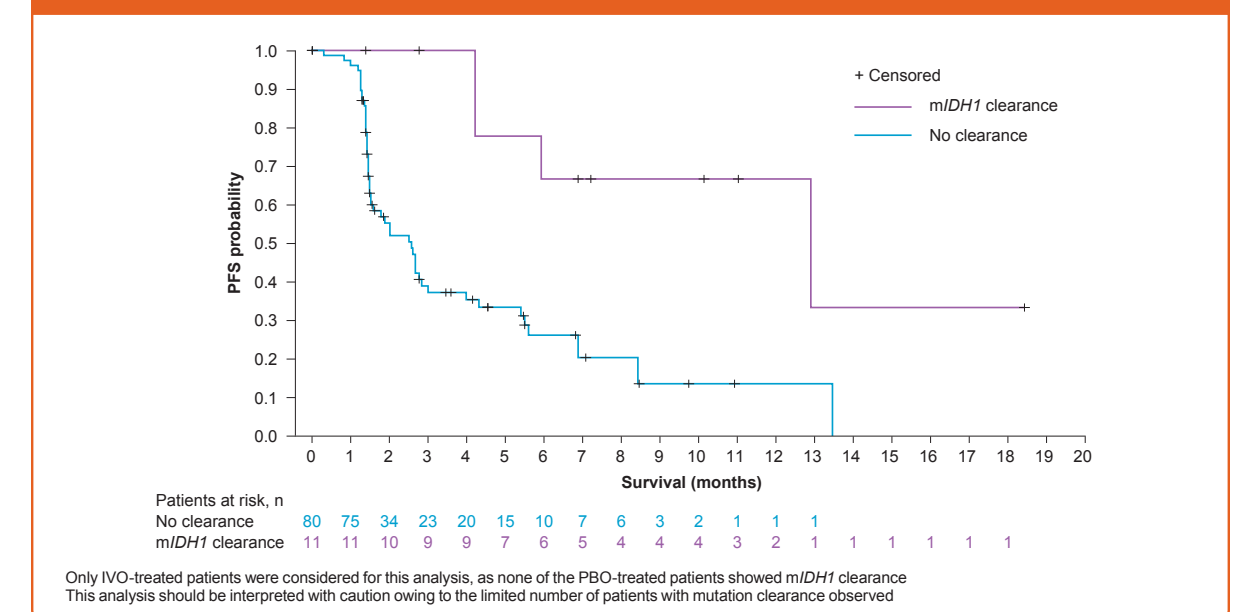


Table 4. Summary of m*IDH1* clearance in plasma

Patients with clearance, n / N (%)	PBO	IVO	
	All	PFS < 2.7 months	PFS ≥ 2.7 months
	0 / 49 (0%)	1 / 55 (1.8%)	10 / 36 (27.8%)

Figure 6. Association between m*IDH1* clearance in plasma and PFS in IVO-treated patients



CONCLUSIONS

- The results obtained from the ClarIDHy study reinforce the feasibility of m*IDH1*-R132 detection in plasma ctDNA from patients with ICC, showing a 92% concordance rate with detection in tumor tissue
- These data support the viability of using liquid biopsy for selecting patients in settings where tissue exhaustion can limit use of conventional methods
- Higher baseline plasma m*IDH1* levels were associated with shorter PFS in patients treated with IVO; no differences were found in the PBO control arm
- m*IDH1* clearance in plasma was observed in a subset of IVO-treated patients and was associated with longer PFS

Acknowledgments

We thank the participating patients and their families.

Disclosures

This study is funded by Agios Pharmaceuticals, Inc. Full author disclosures are available through the ASCO meeting library. Editorial assistance was provided by Vanessa Duca, PhD, Excel Medical Affairs, Fairfield, CT, USA, and supported by Agios.

References

- Dang L et al. Nature 2009;462:739–44.
- Lu C et al. Nature 2012;483:474–8.
- Turcan S et al. Nature 2012;483:479–83.
- Figuerroa ME et al. Cancer Cell 2010;18:553–67.
- Boscoe AN et al. J Gastrointest Oncol 2019;10:751–65.
- Abou-Alfa GK et al. Lancet Oncol 2020 [ePub ahead of print].
- Modi K, Cleary SP. Front Oncol 2018;8:212.
- Goyal L et al. Mol Cancer Ther 2018;17(1 Suppl):A183.
- Goyal L et al. Cancer Discovery 2017;7:252–63.
- Etrich T et al. Sci Rep 2019;9:13261.
- Aguado-Fraile E et al. Cancer Res 2019;79(13 Suppl):Abstr 2275.
- Diehl F et al. Proc Natl Acad Sci U S A 2006;102:16368–73.